Structures of Stealthins A and B, New Free Radical Scavengers of Microbial Origin

Kazuo Shin-ya, Kazuo Furihata,[†] Yoshihiro Teshima, Yoichi Hayakawa and Haruo Seto^{*} Institute of Applied Microbiology, The University of Tokyo, Bunkyo-ku, Tokyo 113, Japan [†]Department of Agricultural Chemistry, The University of Tokyo, Bunkyo-ku, Tokyo 113, Japan.

Summary. Stealthins A and B were isolated as potent radical scavengers from Streptomyces viridochromogenes. They possess an unprecedented benzo[b]fluoren-10-one skeleton as shown in Figure 1.

Over the past decade, many diseases have proven to be caused by oxygen-derived free radicals, such as atherosclerosis, inflammation, Parkinson's disease and ischemic injuries to the central nervous system (CNS) and cardiovascule.¹⁻³ These diseases have been reported to be ameliorated by free radical scavengers such as superoxide dismutase (SOD).⁴

In the course of our screening for free radical scavengers of microbial origin to overcome these diseases, we isolated naphterpin,⁵ antiostatins,⁶ pyridoxatin⁷ and benthocyanins.⁸ Further investigation has resulted in the isolation of potent free radical scavengers stealthins A and B (1 and 2, Fig. 1) from *Streptomyces*



1: $R_1 = OH$, $R_2 = R_3 = R_4 = R_5 = H$ 2: R_1 , $R_2 = O$, $R_3 = R_4 = R_5 = H$ 3: $R_1 = OH$, $R_2 = R_3 = H$, $R_4 = R_5 = CH_3$ 4: R_1 , $R_2 = O$, $R_3 = R_4 = R_5 = CH_3$

Figure 1. Structures of stealthins A (1) and B (2) and their derivatives.

viridochromogenes 2220-SV2. The identification and fermentation of this strain will be reported elsewhere.

Acetone extraction of the mycelium of the producing organism followed by EtOAc extraction gave a crude active material. Silica gel column chromatography using CHCl₃-MeOH (5 : 1) as a solvent system and preparative TLC with CHCl₃-MeOH (10 : 1) gave 1 and 2 as a red powder and a purple powder, respectively. Since they were very unstable under the light, their isolation was quickly carried out in the dark.

They were characterized as follows; 1, UV nm (ɛ) in MeOH 274 (37,600), 470 (sh, 7,500), 504 (11,600), 538 (13,900); MeOH + HCl 269 (36,400), 275 (37,600), 330 (7,000), 430 (5,800), 505 (8,700), 538 (11,000); in MeOH + NaOH 268 (29,500), 275 (30,100), 357 (11,600), 583 (12,100); IR (KBr) 3400, 3220, 3200, 1635 cm⁻¹; and 2, UV nm (c) in MeOH 286 (21,900), 315 (19,200), 415 (6,300), 550 (sh, 11.600), 585 (13,900); in MeOH + HCl 268 (34,000), 275 (35,300), 330 (5,500), 345 (8,100), 415 (4,900), 437 (6,700), 503 (9,800), 535 (12,500); IR (KBr) 3430, 1680, 1630 cm⁻¹. The molecular formulae of 1 and 2 were determined as $C_{18}H_{13}NO_5$ [m/z, (M+H)⁺, 324.0904 ($C_{18}H_{14}NO_5$, + 3.2 mmu error), m-nitrobenzyl alcohol matrix] and C₁₈H₁₁NO₅ [m/z, (M+H)⁺, 322.0756 (C₁₈H₁₂NO₅, + 4.0 mmu error), m-nitrobenzyl alcohol matrix], respectively, from their HR-FAB mass spectra. UV spectral similarity between 1 and 2 suggested the presence of the same chromophore in the both compounds. Surprisingly, however, no signals were observed in the ¹H- and ¹³C-NMR spectra of these compounds presumably due to extensive line broadening.⁹ Since the presence of hydroxy and/or amine functions was indicated by their IR spectral data (3400, 3320 and 3220 cm⁻¹), methylation of 1 was carried out by treatment with CH₃I in the presence of K_2CO_3 . Among several methylated products,¹⁰ the major component, O^5 , N-dimethylstealthin A (3)¹¹ gave nice NMR spectra suitable for structural analysis. The molecular formula of 3, C20H17NO5, was established by HR-FAB mass spectral data [m/z, (M+H)+, 352.1185 (+ 1.0 mmu error)]. Its ¹H- and ¹³C-NMR spectral data revealed the presence of an unprecedented benzo[b]fluoren-10-one structure.

The ¹H-NMR spectral analysis of **3** taken in CDCl₃ identified three separate proton spin systems, a 1,2,3- trisubstituted benzene, an aromatic hydroxy methyl proton (12-H) long-range coupled to two aromatic



Figure 2. NMR analyses of O^5 , N-dimethylstealthin A (3) (in CDCl₃).

methines (1-H and 3-H), which were *meta* coupled each other, and a monomethyl amino residue. The remaining protons were assigned as follows; a methoxy residue (at C-5), a phenolic hydroxy proton (9-OH) hydrogen-bonded to a carbonyl at a *peri* position and a phenolic hydrogen (4-OH).

These separated units were combined to give two ring units (substituted A and D rings) and unassigned sp^2 carbons (106.7 ppm and either 125.1 ppm or 135.3 ppm) as shown in Fig. 2 based on detailed analysis of ${}^{13}C^{-1}H$ long-range couplings observed in the HMBC spectrum of 3. Due to the lack of ${}^{1}H^{-13}C$ long range couplings, C-11a was tentatively assigned to the signal at 135.3 ppm. The two ring units could be combined by NOE observation between the phenolic hydroxy proton at C-4 and methoxy protons at C-5, and the remaining two carbons were incorporated into the ring system by elimination to give the total structure of 3 as shown in Fig. 1. Taking into consideration of the substituents on the adjacent carbons, C-4b and C-10a were assigned to the signals at 125.1 ppm and 106.7 ppm, respectively.

Methylation of 2 gave only a trimethyl derivative (4, $C_{21}H_{17}NO_5$, HRFAB-MS m/z, (M+H)⁺, 364.1172, $C_{21}H_{18}NO_5$ requires 364.1185).¹² Its ¹H- and ¹³C-NMR spectral data proved the presence of an aldehyde group (9.98 ppm and 190.5 ppm, respectively) in 4. Long-range ¹³C-¹H couplings from 1-H and 3-H to the aldehyde carbon implied that 2 was an oxidized derivative of 1 with the aldehyde group at C-12. The very close similarities of the remaining signals between 2 and 4 established the structure of 4 to be $O^{4,5}$, N-trimethylstealthin B.¹³,¹⁴ Thus, the structure of 2 proved to be as shown in Fig. 1.

The benzo[b]fluoren-10-one skeleton present in 1 and 2 is very unique, and as far as we know, this structural unit has never been reported so far. Failure of observation of any signals in the ¹H- and ¹³C-NMR spectra of 1 and 2 is reasonably ascrived to severe signal broadening caused by many tautomeric forms of this unique chromophore including 4-, 5-, 9- and 10-keto forms and 11-imine form.

Stealthins A and B exhibited a potent *in vitro* free radical scavenging activity in a rat liver microsome system with IC₅₀ values 0.04 μ g/ml and 0.07 μ g/ml, respectively. These values are approximately 20-30 times as strong as that of vitamin E. They also inhibited the hemolysis of rat erythrocytes at IC₅₀ being 0.93 μ g/ml (1) and 1.60 μ g/ml (2), while vitamin E showed a potent activity in this system with IC₅₀ value 0.49 μ g/ml. Any methyl derivatives showed no activity in both evaluation system at 100 μ g/ml. Further studies on other biological activities are now under way.

Acknowledgments. This work was supported in part by a grant from Japan Antibiotic Research Association to H.S., and by Grant-In-Aid for JSPS fellows to K.S.

References and Notes:

- (1) Hammond, B.; Kontos, H. A.; Hess, M. L. Can. J. Physiol. Pharmacol., 1985, 63, 173.
- (2) Kontos, H. A. Chem. Biol. Interact. 1989, 72, 229.
- (3) Bolli, R.; Jeroudi, M. O.; Patel, B. S.; DuBose, C. M.; Lai, E. K.; Roberts, R.; McCay, P. B. Proc. Natl. Acad. Sci. U. S. A. 1989, 86, 4695.
- (4) Inoue, M.; Watanabe, N.; Morino, Y.; Tanaka, Y.; Amachi, T.; Sasaki, J. FEBS Lett. 1990, 269, 89.
- (5) Shin-ya, K.; Imai, S.; Furihata, K.; Hayakawa, Y.; Kato, Y.; VanDuyne, G. D.; Clardy, J.; Seto, H. J. Antibiotics 1990, 43, 444.

- (6) Mo, C. -J.; Shin-ya, K.; Furihata, K.; Hayakawa, Y.; Seto, H. J. Antibiotics 1990, 43, 1337.
- (7) Teshima, Y.; Shin-ya, K.; Shimazu, A.; Furihata, K.; Ha, S. C.; Furihata, K.; Hayakawa, Y.; Nagai, K.; Seto, H. J. Antibiotics 1991, 44, 685.
- (8) Shin-ya, K.; Furihata, K.; Hayakawa, Y.; Kato, Y.; Clardy, J.; Seto, H. Tetrahedron Lett. 1991, 32, 943.
- (9) The ¹H- and ¹³C-NMR spectra of stealthins were taken by a JEOL A-500 spectrometer operating at 500 MHz and 125 MHz, respectively. All attempts to observe sharp signals of free stealthins by changing temperatures and solvents were unsuccessful.
- (10) In addition to the dimethyl derivative 3, this methylation reaction gave several methylated derivatives assumed to be O^4 , N-dimethylstealthin A, $O^{4,5,12}$, N, N-pentamethylstealthin A, etc. by NMR spectral analysis. They were, however, not fully characterized.
- UV nm (ε) in MeOH 272 (49,400), 330 (5,000), 468 (10,800), 498 (20,600), 534 (22,500); in MeOH + NaOH 271 (48,000), 468 (9,400), 498 (16,800), 534 (18,300); IR (KBr) 3430, 1620 cm⁻¹. Exhaustive permethylation with Ag₂O caused extensive degradation of partially methyl derivatives, and no expected permethyl derivatives could be obtained.
- (12) Permethylation with Ag₂O caused severe degradation of 4.
- (13) ¹H-NMR (CDCl₃) δ 10.48 (11-NH, bs), 9.98 (12-H), 7.90 (1-H), 7.53 (3-H), 7.41 (7-H, t, J = 8 Hz), 7.38 (6-H, dd, J = 8, 1 Hz), 6.86 (8-H, dd, J = 8, 1 Hz), 4.10 (4-OCH₃, 3H, s), 3.89 (5-OCH₃, 3H, s), 3.64 (11-NCH₃, 3H, d, J = 6 Hz); ¹³C-NMR (CDCl₃) δ 190.5 (C-12), 179.3 (C-10), 162.8 (C-9), 161.1 (C-11), 156.3 (C-4), 147.5 (C-5), 137.0 (C-2), 136.4 (C-5a), 135.6 (C-11a^{*}), 133.4 (C-7), 132.2 (C-4a), 122.4 (C-4b^{*}), 120.1 (C-1), 118.6 (C-9a), 115.8 (C-8), 115.3 (C-6), 115.1 (C-3), 107.8 (C-10a^{*}), 64.4 (5-OCH₃), 56.2 (4-OCH₃), 32.6 (11-NCH₃).

*These carbon assignments may be exchangeable.

(14) Reduction of stealthin B with NaBH₄ or NaBH₃CN to prepare stealthin A was unsuccessful because of decomposition.

(Received in Japan 11 July 1992)